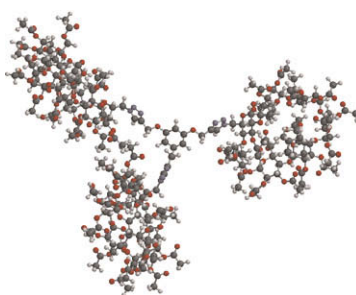


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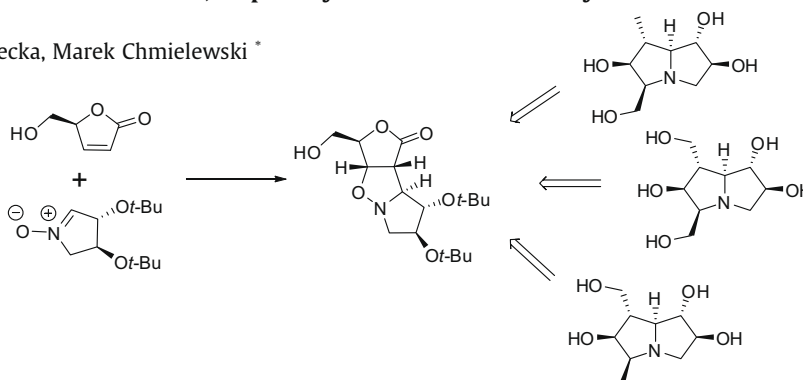
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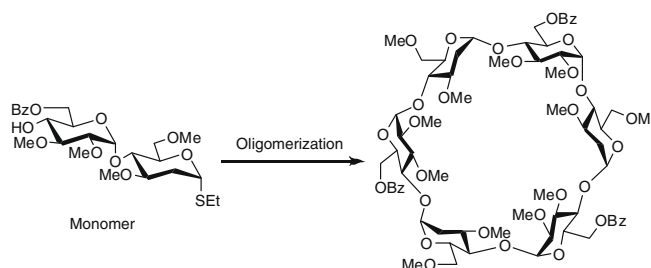
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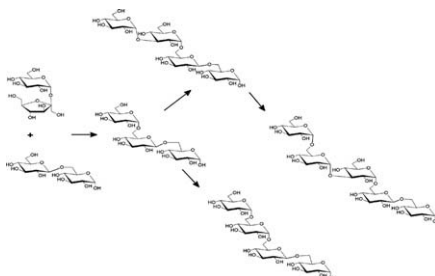
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## Acceptor products of alternansucrase with gentiobiose. Production of novel oligosaccharides for food and feed and elimination of bitterness

pp 187–190

Gregory L. Côté \*

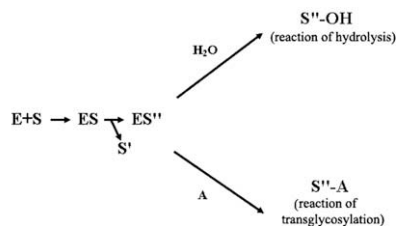


## An endo-(1→3)-β-D-glucanase from the scallop *Chlamys albidus*: catalytic properties, cDNA cloning and secondary-structure characterization

pp 191–197

Svetlana N. Kovalchuk \*, Irina Yu. Bakunina, Yulia V. Burtseva, Viktor I. Emelyanenko, Natalia Yu. Kim, Konstantin V. Guzev, Valeri B. Kozhemyako, Valeri A. Rasskazov, Tatyana N. Zvyagintseva

Endo-(1→3)-β-D-glucanase from the scallop  
*Chlamys albidus* (EC 3.2.1.39) catalyzes two reactions:



## New oligosaccharides prepared by acid hydrolysis of the polysaccharides from *Nerium indicum* Mill and their anti-angiogenesis activities

pp 198–203

Ke Hu, Qin Liu, Shunchun Wang \*, Kan Ding \*

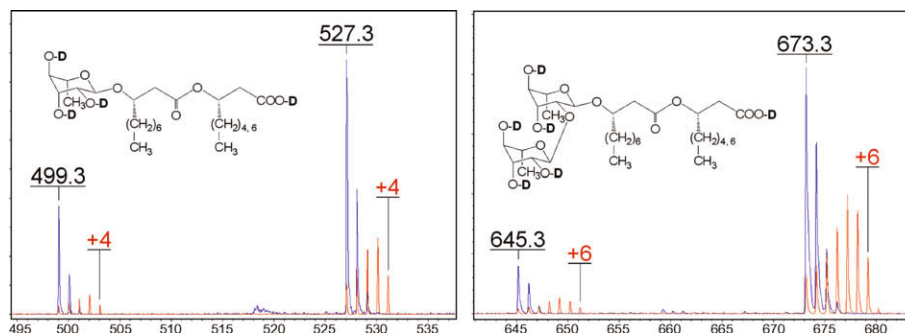
Three oligosaccharides (OJ1–OJ3) were obtained by acid degradation of crude polysaccharides from *Nerium indicum* Mill. The study of chemical and spectroscopic methods demonstrated that they were β-D-Galp-(1→4)-β-D-Galp-(1→4)-D-Galp (OJ1), β-D-Galp-(1→4)-β-D-Galp-(1→3)-D-Galp (OJ2) and another new oligosaccharide, β-D-Galp-(1→4)-β-D-Galp-(1→3)-D-Galp (OJ3). Angiogenesis test showed that OJ2 and OJ3 blocked cell tube formation of HMEC-1.



## MALDI-TOF mass spectrometry of naturally occurring mixtures of monorhamnolipids and dirhamnolipids

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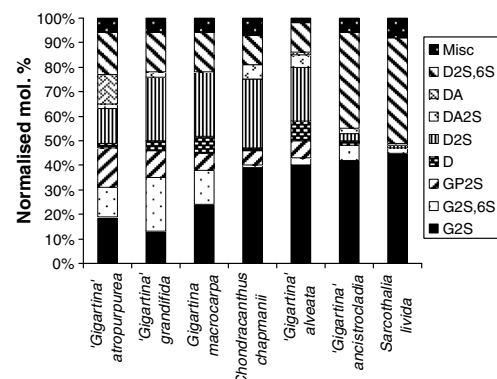
Neil P.J. Price \*, Karen J. Ray, Karl Vermillion, Tsung-Min Kuo



## Chemotaxonomy of New Zealand red algae in the family Gigartinaeae (Rhodophyta) based on galactan structures from the tetrasporophyte life-stage

Ruth Falshaw <sup>\*</sup>, Richard H. Furneaux

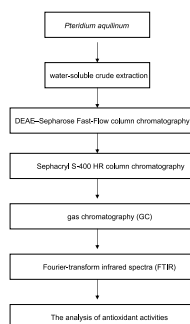
Differences in the structures of the galactans from 15 endemic New Zealand *Gigartina* species indicate seven chemotaxonomically distinct groupings that are aligned to those determined by *rbcL* sequence analysis reported in the literature.



## Antioxidant activity of a water-soluble polysaccharide purified from *Pteridium aquilinum*

Wentao Xu, Fangfang Zhang, YunBo Luo, Liyan Ma, Xiaohong Kou, Kunlun Huang <sup>\*</sup>

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## Synthesis and rheological properties of hydrogels based on amphiphilic alginate-amide derivatives

Frédéric Vallée, Christophe Müller, Alain Durand, Sarah Schimchowitsch, Edith Dellacherie, Christian Kelche, Jean Christophe Cassel, Michèle Leonard <sup>\*</sup>

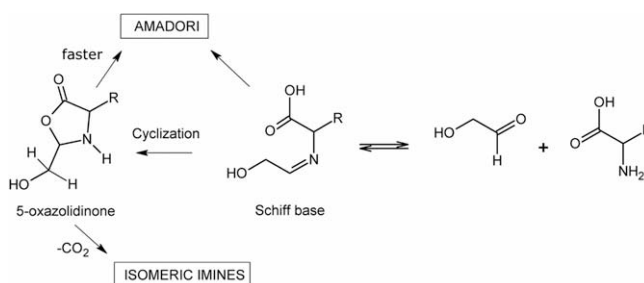
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The properties of amphiphilic ester and amide derivatives of alginate are determined and compared in terms of solubility and stability towards hydrolysis and rheological behaviour.

## FTIR monitoring of oxazolidin-5-one formation and decomposition in a glycolaldehyde–phenylalanine model system by isotope labeling techniques

Fong Lam Chu, Varoujan A. Yaylayan <sup>\*</sup>

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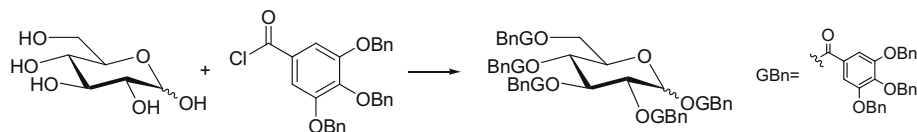


## Notes

**Anomeric selectivity in the synthesis of galloyl esters of D-glucose**

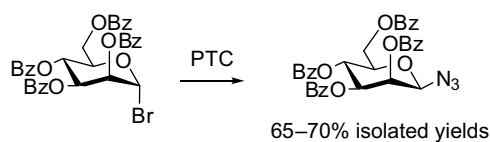
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Robert C. Binkley, Jessica C. Ziepfel, Klaus B. Himmeldirk \*

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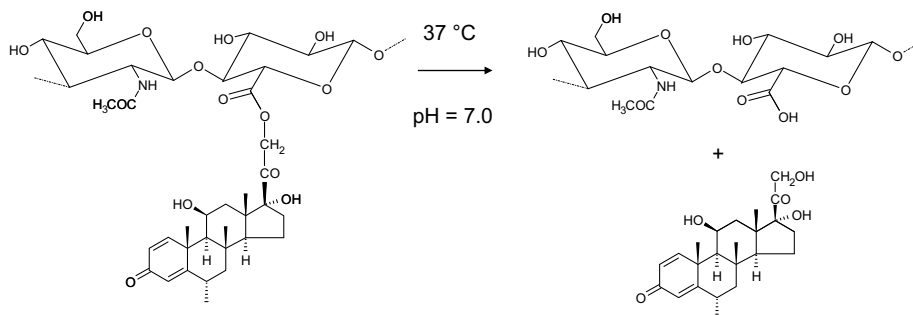
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Polina I. Abronina, Vadim V. Kachala, Leonid O. Kononov \*

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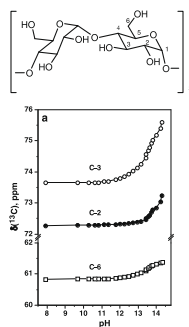
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Anna Taglienti \*, Paolo Sequi, Massimiliano Valentini \*

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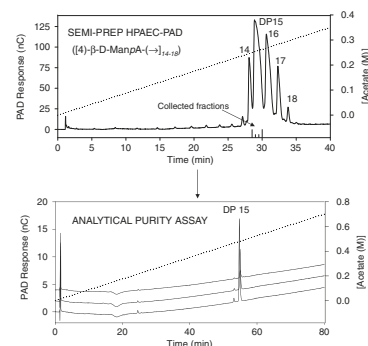
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## Preparation of high purity monodisperse oligosaccharides derived from mannuronan by size-exclusion chromatography followed by semi-preparative high-performance anion-exchange chromatography with pulsed amperometric detection

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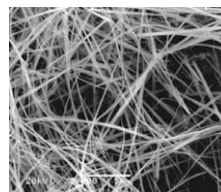
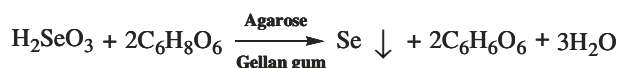
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## Agarose and gellan as morphology-directing agents for the preparation of selenium nanowires in water

pp 260–262

Sanghoo Lee, Sahwan Hong, Baeho Park, Seung R. Paik, Seunho Jung \*



\*Corresponding author

 Supplementary data available via ScienceDirect

### COVER

Shown is a fluorescence image of cell-surface glycans in a 3-day old zebrafish larva. Dierent colors represent glycans biosynthesized at dierent times in development. The glycans were imaged in live zebrafish using a two-step approach termed the bioorthogonal chemical reporter strategy. Embryos were first metabolically labeled with the unnatural monosaccharide *N*-azidoacetylglactosamine, which targets the core position of mucin-type O-glycans; subsequently, the azide-containing glycans were reacted with a cyclooctyne–fluorophore conjugate by copper-free click chemistry, a step that was repeated multiple times to target temporally distinct glycan pools with different fluorophores. This work is the result of a collaboration between the Departments of Chemistry and Molecular and Cell Biology at the University of California, Berkeley [Laughlin, S. T.; Baskin, J. M.; Amacher, S. L.; Bertozzi, C. R. *Science* **2008**, 320, 664].

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Abstracted/Indexed in: Chem. Abstr.: Curr. Contents: Phys., Chem. & Earth Sci. Life Sci. Current Awareness in Bio. Sci. (CABS). Science Citation Index. Full texts are incorporated in CJELSEVIER, a file in the Chemical Journals Online database which is available on STN® International. Also covered in the abstract and citation database SCOPUS®. Full text available on ScienceDirect®



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ISSN 0008-6215